

# FORMATION OF CATHARANTHINE, AKUAMMICINE AND VINDOLINE IN *CATHARANTHUS ROSEUS* SUSPENSION CELLS

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**Key Word Index**—*Catharanthus roseus*; Apocynaceae; plant tissue culture; biosynthesis; indole alkaloids; catharanthine; akuammicine; vindoline.

**Abstract**—Catharanthine and akuammicine, together with ajmalicine and strictosidine, were isolated from a culture strain of *Catharanthus roseus* suspension cells. The biosynthetic capability of the cultured cells to produce akuammicine, catharanthine and vindoline was confirmed by feeding experiments with DL-tryptophan-[3-<sup>14</sup>C] to yield the radioactive alkaloids.

## INTRODUCTION

During the past few years important information about the biosynthesis of *Corynanthe*-type alkaloids has been obtained with cell-free systems of *Catharanthus roseus* cultured cells [1, 2]. However, the hypothetical pathway [3] for the biosynthesis of *Iboga*-, *Aspidosperma*- and *Strychnos*-type alkaloids remains untested since proof for presence of any indole alkaloids beyond the *Corynanthe*-type in the cultured cells is not yet rigorous. There is preliminary evidence for akuammicine (1) [4] and vindoline (2) [5] in *C. roseus* callus based on TLC data, but a note has also appeared to the effect that catharanthine (3), vindoline (2) and dimeric alkaloids are not found in the callus [6]. In this paper we describe evidence for the production of catharanthine (3) (*Iboga*), akuammicine (1) (*Strychnos*) and vindoline (2) (*Aspidosperma*) by a strain of *C. roseus* suspension cultures.

## RESULTS AND DISCUSSION

Callus tissues derived from *C. roseus* seedlings were maintained for more than 1 year on SH medium [7] in

Petri dishes. Distinguishable suspension cultures initiated from callus tissues produced in different Petri dishes have been maintained for a further year as culture strains before screening their alkaloid compositions. One strain (CRW) was found to have an enriched alkaloid composition, including akuammicine (1) and catharanthine (3), as well as ajmalicine (4) and strictosidine (5).

These alkaloids were isolated and identified by means of chromatographic, physicochemical and spectroscopic methods. The content of catharanthine (3) (2 mg from 5.1 kg wet cells) in this strain is similar to that of ajmalicine (4) (1.9 mg from 5.1 kg wet cells). A small amount of akuammicine (1) was also isolated. However, vindoline (2), which is another main indole alkaloid of *C. roseus* intact plants, could not be isolated from any of the culture strains, although a feeding of tryptophan-[<sup>14</sup>C] to the suspension cultures indicated the production of this alkaloid. Since tryptophan is a precursor of the indole alkaloids [8–10], DL-tryptophan-[3-<sup>14</sup>C] was fed to the suspension cultures of the callus, leading to a positive incorporation of the tracer into akuammicine (1), vindoline (2), catharanthine (3) and ajmalicine (4), as shown in Table 1.

Table 1. Incorporation of DL-tryptophan-[3-<sup>14</sup>C] into some indole alkaloids by suspension cultures of *C. roseus* cells

Feeding time (hr)	24	48
Fresh wt of cells (g)	17.8	21.7
Tryptophan fed (dpm)	$5.02 \times 10^7$	$5.02 \times 10^7$
Tryptophan recovered (dpm)	$5.17 \times 10^6$	$4.71 \times 10^6$
Total radioactivity of alkaloids fr. (dpm) [% Incorp.]*	$3.92 \times 10^6$ [7.8]	$4.07 \times 10^6$ [8.1]
Akuammicine (1) isolated (dpm) [% Incorp.]*	$9.49 \times 10^3$ [0.019]	$1.05 \times 10^4$ [0.021]
Vindoline (2) isolated (dpm) [% Incorp.]*	$8.02 \times 10^2$ [0.0016]	$9.14 \times 10^2$ [0.0018]
Catharanthine (3) isolated (dpm) [% Incorp.]*	$7.51 \times 10^4$ [0.15]	$1.02 \times 10^5$ [0.20]
Ajmalicine (4) isolated (dpm) [% Incorp.]*	$9.52 \times 10^4$ [0.19]	$1.33 \times 10^5$ [0.27]

\* No adjustment made for recovered precursor.

The modified capability of this strain to produce complex indole alkaloids has been retained for at least 4 months (8 generations). The present study shows that production of *Strychnos*-, *Aspidosperma*- and *Ipoga*-type alkaloids can be retained (or induced) in some cultured cells. The genetic basis of such a cellular variation for alkaloid biosynthesis remains unclear. However, it is of interest to note that the isolation of this culture strain should lead to the elucidation of the biosynthesis of *Strychnos*-, *Aspidosperma*- and *Ipoga*-type alkaloids in cell-free systems and that it may be possible to obtain variant cell lines producing dimeric alkaloids of the vinblastine type by further selection in the near future.

#### EXPERIMENTAL

**Plant material and culture method.** Callus tissues were induced from seedlings of *C. roseus* (L.) G. Don on SH medium [7] containing 0.8% agar. Subcultures have been maintained on the same medium for more than a year. Suspension cultures were initiated by inoculating callus tissues into liquid SH medium and subcultured in the same medium. Suspension cells were incubated on a rotary shaker (ca 100 rpm) under dim light at ca 27°. For isolation of alkaloids the cultured cells were grown at room temp. in a 14 l. fermentor (New Brunswick Microferm).

**Extraction and isolation of alkaloids.** After 14 days culture, suspension cells (5.1 kg wet wt) were harvested and extracted with MeOH. After removal of solvent, the residue was diluted with K<sub>2</sub>CO<sub>3</sub> soln and then extracted with CHCl<sub>3</sub> to give an oily mass (271 mg). Crude oil was subjected to TLC on Si gel plates (1 mm) using 4 different solvent systems [(i) CHCl<sub>3</sub>-MeOH, 17:3, (ii) petrol-Me<sub>2</sub>CO-diethylamine, 7:2:1, (iii) petrol-CHCl<sub>3</sub>-Me<sub>2</sub>CO, 8:5:4 and (iv) Et<sub>2</sub>O] and HPLC (Waters equipment) using a  $\mu$ C<sub>18</sub>-Bondapak column eluted with MeOH-10 mM K-Pi (pH 7.2) (3:2) at a flow rate of 2 ml/min, to afford catharanthine (**3**) (2 mg) [UV  $\lambda_{\text{max}}^{\text{MeOH}}$  (log  $\epsilon$ ): 291 (3.86), 283 (3.91), 224 (4.55); CD (c 9.6  $\times 10^{-4}$ , EtOH) [ $\theta$ ]<sub>273</sub> +12 000, [ $\theta$ ]<sub>239</sub> -26 200; MS *m/e* (rel. int.): 336 (70, M<sup>+</sup>), 135 (100); <sup>1</sup>H NMR(CDCl<sub>3</sub>):  $\delta$  1.13 (t, 3H, *J* = 7 Hz), 3.77 (s, 3H), 4.79 (s, 1H), 6.26 (*ed*, 1H, *J* = 6 Hz), 7.93 (*bs*, 1H)] and akuammicine (**1**) (125  $\mu$ g) [UV  $\lambda_{\text{max}}^{\text{MeOH}}$ : 328, 296, 225; [ $\alpha$ ]<sub>D</sub><sup>25</sup> -700° (c 0.01, MeOH); MS *m/e* (rel. int.) 322 (10, M<sup>+</sup>), 121 (100); <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  1.57 (d, 3H, *J* = 6 Hz), 3.73 (s, 3H), 5.30 (q, 1H, *J* = 6 Hz)]. In

addition, ajmalicine (**4**) (1.9 mg) [identified with direct comparison of mmp, co-TLC, co-HPLC, UV and IR] and stricoidine (**5**) (4.5 mg) [direct comparison of its co-TLC, co-HPLC and IR] were also isolated.

**Feeding of DL-tryptophan-[3-<sup>14</sup>C] into *C. roseus* cells.** Labeled tryptophan (0.44 mol, 5.02  $\times 10^7$  dpm) dissolved in H<sub>2</sub>O was fed to suspension cells cultured for 1 week prior to feeding. After incubation for 1 or 2 days (see Table 1) at 27° under dim light, the callus was extracted with MeOH, then re-extracted with CHCl<sub>3</sub> in the same manner as above to afford crude alkaloids. After addition of carrier materials (each 10 mg), the crude alkaloids were subjected to 2-D TLC [Si gel; solvent (i) CHCl<sub>3</sub>-MeOH, 17:3 and (ii) Et<sub>2</sub>O] to afford catharanthine, akuammicine, ajmalicine and vindoline, which were recrystallized repeatedly to constant sp. act. Results are summarized in Table 1.

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